

Inducement of Seed Priming with Potassium Nitrate on quality Performance of Chickpea (*Cicer arietinum* L.)

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(Received 02 September 2022, Accepted 28 October, 2022)

(Published by Research Trend, Website: www.researchtrend.net)

ABSTRACT: Seed priming is a pre-sowing treatment which results in a physiological condition that allows seed to germinate more efficient. During subsequent germination, primed seeds exhibit a faster and more synchronized germination and young seedlings are often more vigorous against abiotic stresses than seedlings obtained from unprimed seeds. Priming often involves soaking seed in predetermined amounts of water or limitation of the imbibition time. The imbibition rate may well be somehow controlled by specific salt agents like KNO_3 . The laboratory experiment was carried out in seed testing laboratory, Department of Seed science and Technology, Bidhan Chandra Krishi Viswavidyalaya, Mohanpur, Nadia, West Bengal, India. This study investigated the germination and seedling vigour of chickpea variety ‘Anuradha’ under different KNO_3 concentration and duration with the objective of enhancing the germination potential before sowing. In the present experiment therefore an attempt has been made to study the effects of seed priming with 50 ppm KNO_3 for 6 hrs, 50 ppm KNO_3 for 8 hours, 50 ppm KNO_3 for 10 hours; 100 ppm KNO_3 for 6 hrs, 100 ppm KNO_3 for 8 hrs, 100 ppm KNO_3 for 10 hours; 200 ppm KNO_3 for 6 hours, 200 ppm KNO_3 for 8 hrs; 200 ppm KNO_3 for 10 hrs; 400 ppm KNO_3 for 6 hrs; 400 ppm KNO_3 for 8 hrs; 400 ppm KNO_3 for 10 hrs and dry seeds as control in laboratory condition. From the experiment, it can be concluded that 100 ppm KNO_3 soaking for 8 hrs was the best performer than other priming materials as it was highest performer in germination percentage (96.30), seedling fresh weight (1.84 g), seedling dry weight (0.14g), seedling Vigour Index-I (2013.08), mean germination time (3.15 days), time to 50 % germination (2.45 days) and germination index (40.453).

Keywords: Germination, potassium nitrate, priming, vigour.

INTRODUCTION

Chickpea (*Cicer arietinum* L.) is a self-pollinated crop. It is diploid ($2n=16$) species with genome size $1C=740$ Mbp (Arumuganathan and Earle 1991) and belonging to sub family Papilionaceae of the family Leguminaceae (Poehlman and Sleper 1995). Chickpea is a cool season legume crop and is grown in several countries worldwide as a food source. Seed is the main edible part of the plant and is a rich source of protein (23.3-28.9%), carbohydrates (61.5%), fats (4.5%) and minerals (phosphorus, calcium, magnesium, iron, zinc). It consists of remarkable attributes like wider climatic adaptation, low production cost and having an ability to be applied in crop alternation and atmospheric nitrogen fixation. Seeds undergo deterioration at various levels during storage resulting in decline in a vigour and viability (Bordolui *et al.*, 2015). Seed priming is one of the most important physiological methods which improves the seed performance and provides faster and synchronized germination (Chakraborty and Bordolui 2021). The primed seeds give earlier, more uniform and sometime greater germination and seedling establishment and growth (Bradford, 1986). It is a physiological strategy that involves soaking of seeds in a solution of a specific priming agent followed by

drying of seeds that initiates germination related process. This has been recognized as an important technology to obtain good germination, rapid development and improved yields in some field crops. Moreover, priming is a simple, low-cost and low-risk technology which would be appropriate for all farmers, irrespective of their socio-economic status. The fundamental principle is that sowing of primed seed reduces the time of germination and may allow germination and growth of seedling under adverse soil and environmental conditions (Farooq *et al.*, 2008). Besides better germination and seedling growth, farmers have reported that primed seeds grew more vigorously, flowered earlier and yielded higher (Harris *et al.*, 2001). By seed priming, maintenance of viability, vigour, and increase the storability of seeds is very much essential (Bordolui *et al.*, 2018). The effectiveness of the priming with simple salt solution, perhaps, depends both on the osmotic potential and the chemical nature of the salt species used. It is reported that nitrate containing compounds may function more efficiently than other salts as priming agents. So, our objective to determination of appropriate concentration and duration of KNO_3 is essential for priming of chickpea seed. Keeping the above points, the present

investigation was carried out after seed prime with KNO_3 in different concentrations and durations, and dry seeds as control in laboratory condition on germination, seedling growth and vigour status.

MATERIALS AND METHODS

The laboratory experiment was carried out in seed testing laboratory, Department of Seed Science and Technology, Bidhan Chandra Krishi Viswavidyalaya, Mohanpur, Nadia, West Bengal, India during 2021 following Complete Randomized Design with three replications. Chickpea (Anuradha) was collected from AICRP pulses in Bidhan Chandra Krishi Viswavidyalaya, Mohanpur, Nadia, West Bengal for this investigation.

Seed priming. Seed priming was done with 50 ppm KNO_3 for 6 hrs (T_1); 50 ppm KNO_3 for 8 hours (T_2); 50 ppm KNO_3 for 10 hours (T_3); 100 ppm KNO_3 for 6 hrs (T_4); 100 ppm KNO_3 for 8 hrs (T_5); 100 ppm KNO_3 for 10 hours (T_6); 200 ppm KNO_3 for 6 hours (T_7); 200 ppm KNO_3 for 8 hrs (T_8); 200 ppm KNO_3 for 10 hrs (T_9); 400 ppm KNO_3 for 6 hrs (T_{10}); 400 ppm KNO_3 for 8 hrs (T_{11}); 400 ppm KNO_3 for 10 hrs (T_{12}). Non-primed seeds were the control (T_0).

Germination Parameters

Time to 50% germination. According to Association of Official Seed Analysis (1983) number of seeds germinated was recorded in daily basis. The time to obtain 50% germination (T_{50}) was calculated according

to the following formulae given by Coolbear *et al.* (1984) which was modified by Farooq *et al.* (2005).

$$T_{50} = t_i + \frac{\left(\frac{N}{2} - n_i\right) (t_j - t_i)}{(n_j - n_i)}$$

Where, N stands for final number of germination and n_i , n_j are cumulative number of seeds germinated by adjacent counts at times t_i and t_j when $n_i < N/2 < n_j$.

Mean germination time (MGT)

Mean germination time (MGT) was calculated with the following equation suggested by Ellis and Roberts (1981).

$$MGT = \frac{\sum D_n}{\sum n}$$

Where n indicates the number of seeds germinated on day D, and D is the number of days counted from the beginning of germination.

Germination percentage. Germination percentage (G) was calculated as:

$$\frac{\text{Number of normal seedlings produced}}{\text{Total number of seeds used}} \times 100$$

Where, X is the number of normal seedlings produced and Y denotes total number of seeds taken for germination (ISTA, 1996). It is expressed in percentage.

Germination index (GI). Germination index (GI) was calculated as described in the Association of Official Seed Analysts (AOSA, 1990) as the following formulae:

$$GI = \frac{\text{No. of germinated seeds}}{\text{Day of fi count}} + \dots + \frac{\text{No. of germinated seeds}}{\text{Day of last count}}$$

Germination Energy. According to Ruan *et al.* (2002) energy of germination (GE) should be recorded at 4th day after planting. It is the percentage of germinating seeds 4 days after planting relative to the total number of seeds tested.

Seedling parameters. Root lengths and shoot lengths of ten seedlings were measured at 8 days after germination by glass plate method in the laboratory with the help of a scale and graph paper and average was made out, expressed in centimetre (cm). Fresh weight of ten seedlings was measured with the help of a digital balance. Then seedlings were dried at 60-70 °C for two hours in hot air oven and weighed in a digital balance. Both seedling fresh weight and dry weight are expressed in gram (g).

Vigour index. Vigour index (VI) was calculated by using the formula suggested by Abdul Baki and Anderson (1973): $VI = G \times L$

Where, 'G' indicates germination percentage and 'L' denotes average seedling length (cm)

RESULT AND DISCUSSION

Germination Index. Germination Index significantly varied due to priming with different duration and concentration of KNO_3 . Lowest Germination index was observed in control (23.060) preceded by T_{12} , T_7 and T_6 . While, T_5 (40.453) showed the highest germination index followed by T_2 , T_4 and T_1 . But, non-significant

difference was observed in between T_6 , T_7 and T_{10} ; T_3 , T_9 and T_{11} ; T_6 and T_9 . Similar type of result was observed by Patil *et al.* (2018); Ray and Bordolui (2022a).

Root length (cm). Significant difference was observed in root length for potassium nitrate priming. Maximum seedling root length was observed for T_8 (6.26 cm) followed by T_3 and T_9 respectively, while it was minimum for T_0 (4.00 cm) (Table 1). Although T_1 , T_4 and T_5 ; T_3 , T_8 and T_9 ; T_6 and T_{11} ; T_0 and T_{12} showed non-significant difference among themselves. Root and shoot length increased in seeds due to priming as compared to non-primed seeds reported by Demir and Oztokat (2003); Ray and Bordolui (2022b).

Shoot length (cm). Considering shoot length, the longest seedling shoot length was recorded for T_1 (17.44 cm) followed by T_5 and T_6 while shortest shoot length was observed in Control (7.56 cm) preceded by T_3 and T_{10} . Significant difference was noted for shoot length in overall though non-significant difference was observed in between T_2 , T_9 and T_{12} ; T_6 and T_{11} ; T_7 and T_{10} . The result corroborates the findings of Chang-Zheng *et al.* (2002) who reported that rice seed treated with mixed salt solution germinated significantly more rapidly than unprimed seed.

Fresh weight (g). Significant difference was observed in fresh weight after potassium nitrate priming. Highest seedling fresh weight was observed for T_5 (1.84 g)

followed by T₁₀ and T₃ while lowest was noted in T₇ (0.85 g) preceded by T₀ and T₂ respectively. But non-significant difference was noticed in between T₆ and T₁₁; T₁₁ and T₉; T₄ and T₁₂; T₅ and T₈.

Dry weight (g). In case of dry weight, it was significantly varied due to priming with different duration and concentration of KNO₃. Maximum seedling dry weight was noticed for T₅ (0.14 g) followed by T₁₁ and T₆ respectively while minimum

was noticed for control (0.07) preceded by T₂ and T₉ respectively. Although non-significant difference was observed in between T₀, T₂ and T₇; T₁, T₃, T₄, T₆, T₁₀ and T₁₂; T₁₀ and T₁₁; T₆ and T₈. Mohammadi (2009) on soybean (*Glycine max* L.) in field and laboratorial studies found that seed primed with potassium nitrate increased germination percentage (GP), seedling fresh weight and seedling dry weight as compared to control.

Table 1: Effect of priming on germination index, root length, shoot length, fresh weight and dry weight of chickpea.

Treatment	Germination Index	Root length (cm)	Shoot length (cm)	Fresh weight (g)	Dry weight (g)
T ₀	23.060	4.00	7.56	0.91	0.07
T ₁	29.897	4.63	17.44	1.23	0.11
T ₂	37.917	5.22	11.87	0.93	0.08
T ₃	27.800	6.17	9.58	1.57	0.10
T ₄	31.163	4.61	14.12	1.36	0.10
T ₅	40.453	4.65	16.26	1.84	0.14
T ₆	26.740	5.83	15.44	1.54	0.12
T ₇	26.207	4.51	11.06	0.85	0.08
T ₈	29.633	6.26	13.93	1.84	0.13
T ₉	27.480	6.07	12.29	1.22	0.10
T ₁₀	26.550	4.51	10.90	1.62	0.12
T ₁₁	27.617	5.57	15.08	1.54	0.13
T ₁₂	24.677	4.06	12.29	1.35	0.10
SEm (±)	0.254	0.140	0.169	0.059	0.01
LSD (0.05)	0.744	0.408	0.495	0.172	0.02

Note: T₀ = Control, T₁ = 50 ppm KNO₃ for 6 hrs, T₂ = 50 ppm KNO₃ for 8 hrs, T₃ = 50 ppm KNO₃ for 10 hrs, T₄ = 100 ppm KNO₃ for 6 hrs, T₅ = 100 ppm KNO₃ for 8 hrs, T₆ = 100 ppm KNO₃ for 10 hrs T₇ = 200 ppm KNO₃ for 6 hrs, T₈ = 200 ppm KNO₃ for 8 hrs, T₉ = 200 ppm KNO₃ for 10 hrs, T₁₀ = 400 ppm KNO₃ for 6 hrs, T₁₁ = 400 ppm KNO₃ for 8 hrs, T₁₂ = 400 ppm KNO₃ for 10 hrs.

Germination Percentage. Significant difference was observed in germination percentage. Among the priming treatments, with different duration and concentration of KNO₃, T₅ (96.30 %) recorded highest germination percentage followed by T₂ and T₈. While lowest germination percentage was recorded for T₀ (85.83) preceded by T₃, T₁₀ and T₆ respectively. But, non-significant difference was observed in between T₁, T₄, T₃, T₆, T₁₀, T₁₁ and T₁₂; T₇, T₈, T₉ and T₁₁. This result is in agreement with Mohammadi and Amiri (2010) who reported that seed priming with nitrate solutions led to improved germination rate.

Vigour Index. Considering vigour index, maximum value was calculated for T₅ (2013.08) followed by T₁ and T₈ respectively; minimum vigour index was noted for T₀ (992.18) preceded by T₁₀ and T₃. Although, vigour index was significantly varied, but some non-significant difference was also noticed in between T₁ and T₅; T₃, T₇, T₁₀ and T₁₂; T₂, T₄ and T₉; T₆, T₈ and T₁₁. Mohammadi (2009) reported similar kind of observation that soaking of soybean (*Glycine max*) seed in potassium nitrate had the best effect on germination and seed vigour with late sowing. Ray and Bordolui (2020)

Time to 50 % germination (days). Significant responses were noticed in the priming treatment with different duration and concentration of potassium nitrate under laboratory condition. Minimum time to 50% germination was recorded in T₅ (2.45 days) followed by T₄ and T₃. While, maximum time to 50%

germination was observed for T₀ (3.55 days) preceded by T₁₂ and T₇. Although, time to 50% germination was significantly varied, but some non significant difference was also noticed in between T₁, T₃, T₄, T₈ and T₁₁; T₂, T₆, T₈, T₉, T₁₀ and T₁₁. Dezfuli *et al.* (2008) observed similar type of result in maize.

Mean germination time (days). Considering Mean germination time T₅ (3.15 days) had the shortest mean germination time preceded by T₂ and T₁. While maximum mean germination time was noticed in T₀ (4.40 days) followed by T₁₂. Though significant difference was observed in mean germination time but some non-significant difference was also noticed in between T₆, T₇, T₉, T₁₀ and T₁₁; T₇, T₈, T₉, T₁₁ and T₁₂; T₁, T₂, T₄ and T₆; T₃ and T₄. Similar kind of observation was noted by Vadez *et al.* (1996) in bean.

Germination energy (%). Significant difference was observed in germination energy for potassium nitrate. The maximum energy of germination was recorded in T₂ (76.09) followed by T₁₁, T₄ and T₆ while it was minimum for T₀ (51.57) preceded by T₇ and T₁₂. Seed priming treatments enhanced the energy of germination over that of untreated seeds and maximum energy of germination was recorded with hydro-priming in rice (Mahajan *et al.*, 2011). Low vigour seeds of hybrid sunflower showed significant decrease in mean germination time and increase in germination index as well as germination energy over non-primed low vigour seeds after priming with KH₂PO₄ (Kausar *et al.*, 2009).

Table 2: Effect of priming on germination percentage, vigour index, Time to 50 % germination, Mean germination time and Germination energy of chickpea.

Treatment	Germination Percentage (Tr value)	Vigour Index	Time to 50 % germination (days)	Mean germination time (days)	Germination energy (%) (Tr value)
T ₀	85.83 (67.86)	992.18	3.55	4.40	51.57 (45.88)
T ₁	89.33 (70.93)	1971.34	2.63	3.45	60.16 (50.84)
T ₂	96.07 (78.62)	1641.81	2.88	3.40	76.09 (60.70)
T ₃	88.57 (70.22)	1394.51	2.55	3.73	59.28 (50.33)
T ₄	89.40 (70.99)	1674.53	2.54	3.46	68.84 (56.05)
T ₅	96.30 (78.93)	2013.08	2.45	3.15	63.42 (52.76)
T ₆	88.90 (70.54)	1890.17	2.83	3.66	68.04 (55.56)
T ₇	92.80 (74.49)	1444.27	3.04	3.88	57.60 (49.35)
T ₈	94.24 (76.12)	1902.20	2.74	3.96	65.60 (54.07)
T ₉	92.47 (74.07)	1697.07	2.96	3.87	59.18 (50.27)
T ₁₀	88.73 (70.40)	1367.15	2.82	3.60	59.73 (50.59)
T ₁₁	91.03 (72.60)	1880.57	2.79	3.69	69.37 (56.37)
T ₁₂	89.13 (70.73)	1457.35	3.34	3.94	57.69 (49.40)
SEm (±)	0.774	23.964	0.066	0.095	0.298
LSD (0.05)	2.263	70.048	0.193	0.279	0.872

Note: T₀ = Control, T₁ = 50 ppm KNO₃ for 6 hrs, T₂ = 50 ppm KNO₃ for 8 hrs, T₃ = 50 ppm KNO₃ for 10 hrs, T₄ = 100 ppm KNO₃ for 6 hrs, T₅ = 100 ppm KNO₃ for 8 hrs, T₆ = 100 ppm KNO₃ for 10 hrs, T₇ = 200 ppm KNO₃ for 6 hrs, T₈ = 200 ppm KNO₃ for 8 hrs, T₉ = 200 ppm KNO₃ for 10 hrs, T₁₀ = 400 ppm KNO₃ for 6 hrs, T₁₁ = 400 ppm KNO₃ for 8 hrs, T₁₂ = 400 ppm KNO₃ for 10 hrs.

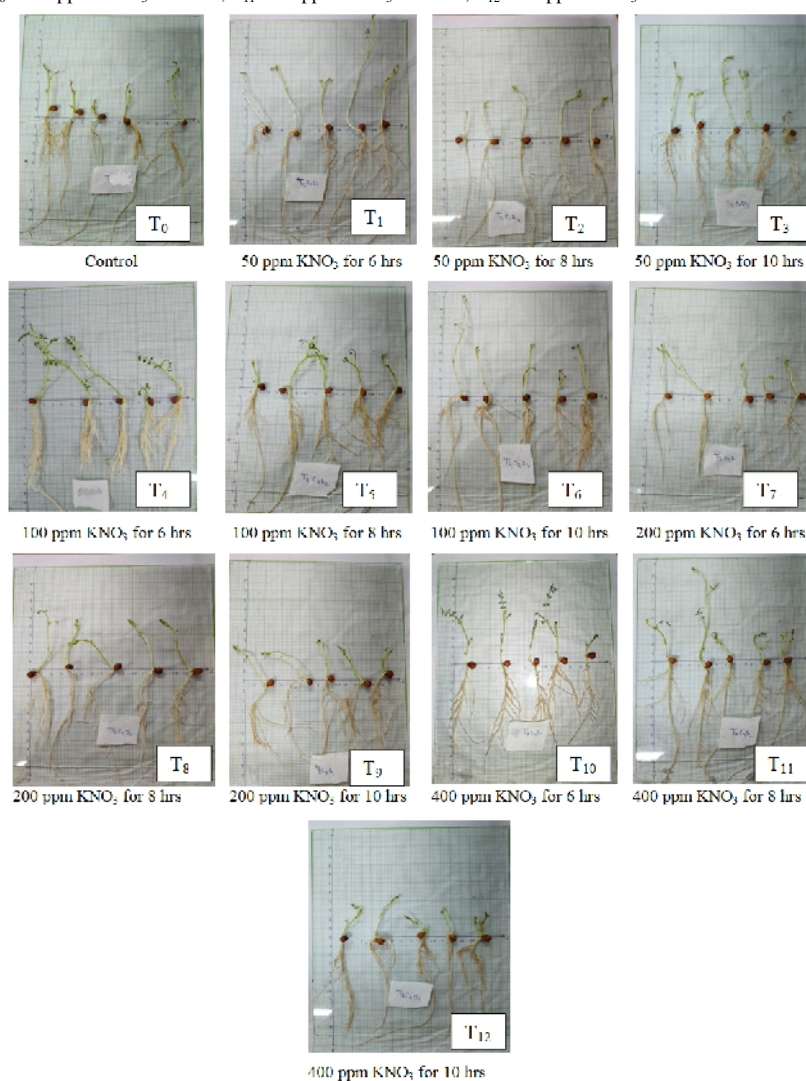


Fig. 1. Evaluation of seedling vigour under laboratory condition

CONCLUSION

Seeds of chickpea were treated with various concentration and duration of potassium nitrate recorded higher seed quality parameters compared to control. Seeds treated with KNO₃ 100 ppm observed the significant higher than other priming concentrations and durations. It can be concluded that among all the treatments, KNO₃ 100 ppm for soaking 8 hrs showed significant performance for seed quality parameter like germination percentage (96.30), seedling fresh weight (1.84 g), seedling dry weight (0.14g), seedling vigour Index-I (2013.08), mean germination time (3.15 days), time to 50 % germination (2.45 days) and germination index (40.453). Therefore, as pre-sowing treatment KNO₃ 100 ppm with a duration of 8 hrs is recommended for treating chickpea seed for better seedling establishment.

FUTURE SCOPE

There is a scope to study the priming effect of other priming materials in chickpea.

Acknowledgement. Authors are thankful to Department of Seed Science and Technology, Bidhan Chandra Krishi Viswavidyalaya, Mohanpur, 741 252, Nadia, West Bengal.

Conflict of Interests. None.

REFERENCES

- Abdul Baki, A. A. and Anderson, J. D. (1973). Vigour determination in soybean seed by multiple criteria I. *Crop science*, 13(6), 630-633.
- Arumuganathan, K. and Earle, E. D. (1991). Nuclear DNA content of some important plant species. *Plant Molecular Biology Reporter*, 9(3), 208-218.
- Association of Official Seed Analysis (1990). Rules for testing seeds. *Journal Seed Technology*, 12, 1112.
- Bordolui, S.K., Chattopadhyay, P. and Basu, A. K. (2018). Evaluation of some small seeded aromatic indigenous genotypes for commercial utilization as high value rice. *International Journal of Minor Fruits, Medicinal and Aromatic Plants*, 4(1), 40-43.
- Bordolui, S. K., Sadhukhan, R. and Chattopadhyay, P. (2015). Participatory evaluation of some folk rice genotypes. *Journal Crop and Weed*, 11(2), 59-62.
- Bradford, K. J. (1986). Manipulation of seed water relations via osmotic priming to improve germination under stress conditions. *Horticulture Science*, 21, 1105-1112.
- Chakraborty, A. and Bordolui, S. K. (2021). Impact of Seed Priming with Ag-Nanoparticle and GA₃ on Germination and Vigour in Green gram. *Int. J. Curr. Microbiol. App. Sci*, 10(03), 1499-1506.
- Chang-Zheng, H., Jin, H., Zhi-yu, Z., Song-Lin, R. and Wen-Jian, S. (2002). Effect of seed priming with mixed-salt solution on germination and physiological characteristics of seedling in rice (*Oryza sativa* L.) under stress conditions. *Journal of Zhejiang University (Agriculture & Life Sciences)*, 28, 175-178.
- Coolbear, P., Francis, A. and Grierson, D. (1984). The effect of low temperature pre-sowing treatment under the germination performance and membrane integrity of artificially aged tomato seeds. *Journal Exp. Botany*, 35, 1609-1617.
- Demir, I. and Oztokat, C. (2003). Effect of salt priming on germination and seedling growth at low temperatures in water melon seeds during development. *Seed Science Technology*, 31, 765-770.
- Dezfuli, P. M., Zadeh, F. S. and Janmohammadi, M. (2008). Influence of Priming Techniques on Seed Germination Behaviour of Maize Inbred Lines (*Zea mays* L.). *ARP Journal of Agricultural and Biological Science*, 3(3), 22-25.
- Ellis, R. A. and Roberts, E. H. (1981). The quantification of ageing and survival in orthodox seeds. *Seed Sci. Technol*, 9, 373-409.
- Farooq, M., Basra, S. M. A., Rehman, H. and Saleem, B. A. (2008). Seed priming enhances the performance of late sown wheat (*Triticum aestivum* L.) by improving the chilling tolerance. *Journal of Agronomy and Crop Sciences*, 194, 55-60.
- Farooq, M., Basra, S. M. A., Hafeez, K. and Ahmad, N. (2005). Thermal hardening: A new seed vigour enhancement tool in rice. *Acta Botanica Sinica*, 47, 187-192.
- Harris, D., Raghuvanshi, B. S., Gangwar, J. S., Singh, S. C., Joshi, K. D. and Rashid, A. (2001). Participatory evaluation by farmers of on-farm seed priming in wheat in India, Nepal, and Pakistan. *Experimental Agriculture*, 37, 403-415.
- ISTA (1996). International Rules of Seed Testing. *Seed Science and Technology*, 24, 1-86.
- Kausar, M., Mahmood, T., Basra, S. M. A. and Arshad, M. (2009). Invigoration of Low Vigour Sunflower Hybrids by Seed Priming. *International Journal Agricultural Biology*, 11, 521-528.
- Mahajan, G., Sarlach, R. S., Japinder, S. and Gill, M. S. (2011). Seed Priming Effects on Germination, Growth and Yield of Dry Direct-Seeded Rice. *Journal of Crop Improvement*, 25(4), 409-417.
- Mohammadi, G. R. (2009). The effect of seed priming on plant traits of late-spring seeded soybean (*Glycine max* L.). *Am. Eur. J. Agric. Environ. Sci.*, 5, 322-326.
- Mohammadi, G. R. and Amiri, F. (2010). The effect of priming on seed performance of canola (*Brassica napus* L.) under drought stress. *Am. Eur. J. Agric. Environ. Sci.*, 9, 202-207.
- Patil, K., Ravat, L., Trivedi, V., Hirpara, A. and Sasidharan, N. (2018). Effect of seed priming treatment in chickpea (*Cicer arietinum* L.). *International Journal of Chemical Studies*, 6(4), 1064-1069.
- Poehlman, J. M. and Sleper, D. A. (1995). Breeding field crops. 3rd ed. Iowa state of university press, Iowa 50014, USA.
- Ray, J. and Bordolui, S.K. (2020). Effect of GA₃ on Marigold Seed Production in Gangetic Alluvial Zone. *Journal of Crop and Weed*, 16(1), 120-126.
- Ray, J. and Bordolui, S. K. (2022a). Effect of Seed Priming as Pre-Treatment Factors on Germination and Seedling Vigour of Tomato. *International Journal of Plant & Soil Science*, 34(20), 302-311.
- Ray, J. and Bordolui, S. K. (2022b). Seed quality deterioration of tomato during storage: Effect of storing containers and condition. *Biological Forum-An International Journal*, 14(2), 137-142.
- Ruan, S., Xue, Q. and Tylkowska, K. (2002). Effects of seed priming on germination and health of rice (*Oryza sativa* L.) seeds. *Seed Science Technology*, 30, 451-458.
- Vadez, V., Rodier, F., Payre, H. and Drevon, J. J. (1996). Nodule permeability to O₂ and nitrogenase linked respiration in bean landraces varying in the tolerance of N₂ fixation to P deficiency. *Plant Physiology Biochemistry*, 34, 871-878.

How to cite this article: Anish Choudhury and Sanjoy Kumar Bordolui (2022). Inducement of Seed Priming with Potassium Nitrate on quality Performance of Chickpea (*Cicer arietinum* L.). *Biological Forum – An International Journal*, 14(4): 779-783.